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MYOPORONE AND RELATED KETO ALCOHOLS FROM STRESSED SWEET POTATOES

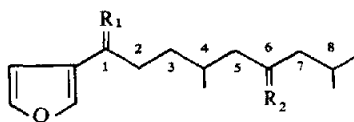
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Key Word Index—*Ipomoea batatas*; Convolvulaceae; sesquiterpene; stress metabolite.

Our investigation of stress metabolites of the sweet potato has led to the isolation of myoporone (**1**) and two related keto alcohols (**2** and **3**) from this plant. The compounds were obtained from a methanol-chloroform extract of mercuric chloride treated sweet potatoes by a combination of column chromatography on Si gel and HPLC. By this method 39 mg of **1**, 109 mg of **2** and 53 mg of **3** were isolated from each kg of treated sweet potatoes.



- 1 $R_1 = R_2 = O$
 2 $R_1 = O$; $R_2 = H, OH$
 3 $R_1 = H, OH$; $R_2 = O$

Myoporone was initially isolated from Si gel chromatography as a mixture with ipomeamarone. The mixture was separated by HPLC on a reversed phase column. Compound **1** [$\alpha_D^{25} -8.5^\circ$, $c = 1.54$, MeOH] was identified by comparing its 1H NMR and mass spectra to those reported by Blackburne *et al.* [1].

The isomeric keto alcohols were isolated from the Si gel column as a mixture with 7-hydroxymyoporone [2]. This mixture was also resolved by HPLC on a reversed phase column. Compound **2** contained a keto-furan moiety as evidenced by the UV λ_{max} (MeOH) 251 nm ($\epsilon = 3000$), IR ν_{max} (neat) 1670 cm^{-1} , and the MS fragments at m/e 95 and 110 corresponding to cleavage of C-1, C-2 and McLafferty cleavage of C-2, C-3. The broad absorption from 3700 to 3100 cm^{-1} in the IR verified the presence of a hydroxyl group. The MS fragment at m/e 195 is consistent with placing the hydroxyl group at

C-6. The 1H NMR spectrum of the compound is consistent with structure **2**: δ 0.91 and 0.93 (9H, superimposed *d*, CH_3), 1.34 (4H, *m*, 5- and 7- CH_2), 1.45–2.2 (5H, *m*), 2.80 (2H, *t*, $J = 7\text{ Hz}$, 2- CH_2), 3.88 (1H, *m*, 6-CH), 6.80 (1H, *m*, 4-furyl), 7.46 (1H, *m*, 5-furyl) and 8.07 (1H, *m*, 2-furyl). The product of oxidation of **2** with pyridinium had chlorochromate [3] 1H NMR and mass spectra identical to those of **1**.

Unlike most of the furanosesquiterpenoid stress metabolites from sweet potatoes, C-1 in compound **3** ($[\alpha_D^{25} -6.4^\circ$, $c = 1.34$, MeOH) is not in the ketone oxidation state. This is obvious from the coincidence of signals in the 1H NMR for the 2- and 5-furyl protons, as is seen in the case of 1-ipomeanol and 1,4-ipomeadiol [4], and from the major fragment in the MS at m/e 97. The carbonyl absorption at 1705 cm^{-1} in the IR spectrum indicates that the carbonyl group is unconjugated. The fragment in the MS at m/e 195 resulting from cleavage at C-6, C-7 and the fragment at m/e 100 resulting from McLafferty cleavage of C-4, C-5 places the carbonyl group at C-6. The 1H NMR of **3** is consistent with the proposed structure: δ 0.90 (9H, *d*, $J = 7\text{ Hz}$, CH_3), 1.34 (2H, four line *m*, 3- CH_2), 1.5–2.2 (CH, *m*), 2.26 (4H, *m*, 5- and 7- CH_2), 4.66 (1H, *t*, $J = 7\text{ Hz}$, 1-CH), 6.42 (1H, *m*, 4-furyl) and 7.42 (2H, *m*, 2- and 5-furyl). Oxidation of **3** with pyridinium chlorochromate [3] gave **1** as the major product.

Myoporone has been isolated from *Myoporum bonitoides* A. Gray [5], from other *Myoporum* species and *Eremophila* species [1], and from *Eumophia sericea* and *E. prostata* [6]. Thus, myoporone joins epingaione [7] (ipomeamarone [8]), dehydroepingaione [7] (dehydroipomeamarone [9]), and athanagrandonone [10] (4-hydroxymyoporone [11]) as furanosesquiterpenes which are stress metabolites of the sweet potato and are also normal secondary metabolites of other plants.

Toxicity studies on compound **2** indicate that it is

hepatotoxic, 2–3 times more potent than ipomeamarone. A detailed report of the toxicity of **2** is forthcoming.

EXPERIMENTAL

NMR spectra were obtained at 100 MHz in CDCl_3 . MS were run via direct probe inlet at 70 eV. IR spectra were obtained as neat samples. HPLC was carried on a Waters Associates instrument using two 30 cm C-18 μ bondapak columns. Column chromatography was carried out using the so-called short column technique of Hunt and Ribby [12].

Sweet potato roots, cut into 0.5 cm slices, were dipped into a 1% soln of HgCl_2 and placed in covered pans for 1 week. At this time the slices were homogenized with MeOH and CHCl_3 . The CHCl_3 extract was dried (MgSO_4) and coned *in vacuo*. The residue was chromatographed on a 5×7 cm column of Merck Sigel G (Type 60) for TLC. For the isolation of myoporone, 1:5 EtOAc–hexane was used as the eluent; for isolation of **2** and **3**, 1:1 EtOAc–hexane was used.

Myoporone was isolated using the C-18 column with 2:1 MeOH– H_2O at a flow rate of 1.7 ml/min as the eluent. Compounds **2** and **3** were isolated by HPLC using the C-18 column with 3:2 MeOH– H_2O at 1.7 ml/min as the eluent. MS of **2**: m/e (rel. int.) 252 [M^+] (3), 195 [cleavage at C-6, C-7] (4), 177 [195 – H_2O] (9), 166 [cleavage at C-5, C-6] (11), 149 [cleavage at C-4, C-5] (11), 123 [cleavage at 3-C, C-4] (20), 110 [McLafferty at C-2, C-3] (64) and 95 [cleavage at C-1, C-2] (100). MS of **3**: m/e (rel. int.) 252 [M^+] (91), 234 [$\text{M}^+ - \text{H}_2\text{O}$] (8), 195 [cleavage at C-6, C-7] (11), 177 [195 – H_2O] (8), 127 [cleavage at G3,

C-4] (100), 100 (McLafferty at C-4, C-5] (32) and 97 [cleavage at C-1, C-2] (83).

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ISOLATION AND IDENTIFICATION OF PHYTUBERIN FROM *NICOTIANA TABACUM* PREVIOUSLY INFILTRATED WITH AN INCOMPATIBLE BACTERIUM*

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Key Word Index—*Nicotiana tabacum*; Solanaceae; incompatible bacteria; sesquiterpene; phytuberin; stress metabolite.

Twenty structurally-related sesquiterpenes have been isolated and characterized from Solanaceous species [1, 2]. Three of these compounds, rishitin, lubimin and capsidiol, have been found in more than one species after stress. To date, only capsidiol has been shown to be a stress metabolite in *Nicotiana tabacum* L. [3, 4]. In this paper, we report the isolation of phytuberin, a sesquiterpene previously only reported in potato [5, 6], from tobacco leaf tissue infiltrated with the bacterium *Pseudomonas lachrymans*, a nonpathogen of tobacco.

The presence of phytuberin in extracts from infected

tobacco tissue was first suggested by TLC analysis which revealed a compound that had the same R_f and produced the same characteristic color as authentic phytuberin with the vanillin– H_2SO_4 reagent [5]. The unknown was determined to be phytuberin by direct comparison of the unknown with an authentic sample [5], by TLC, GLC, chemical ionization MS and IR. Phytuberin was not detected in healthy leaf tissue.

EXPERIMENTAL

Leaves of 6- to 10-week-old *Nicotiana tabacum* L., cv KY 16, were infiltrated with H_2O or a suspension of the incompatible bacterium *Pseudomonas lachrymans* (10^8 cells/ml) prepared from 24-hour-old cultures. The leaves which were infiltrated

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